



MEMORANDUM REPORT ARLCD-MR-77006

FEASIBILITY OF USING FLUORESCENT MATERIALS IN PRODUCT ASSURANCE APPLICATIONS AND FOR LOCATING ADHESIVE BOND FRACTURES

CAROLYN A. L. WESTERDAHL
J. RICHARD HALL

MARCH 1978



US ARMY ARMAMENT RESEARCH AND DEVELOPMENT COMMAND

LARGE CALIBER

WEAPON SYSTEMS LABORATORY

DOVER. NEW JERSEY

APPROVED FOR PUBLIC RELEASE: DISTRIBUTION UNLIMITED.

The findings in this report are not to be construed as an official Department of the Army position.

DISPOSITION

Destroy this report when no longer needed. Do not return to the originator.

The citation in this report of the names of commercial firms or commercially available products or services does not constitute official endorsement or approval of such commercial firms, products, or services by the US Government.

SECURITY CLASSIFICATION OF THIS PAGE (When Dete Entered)

REPORT DOCUMENTATION PAG	READ INST BEFORE COMP	
1. REPORT NUMBER 2. GO Memorandum Report ARLCD-MR-77006	T ACCESSION NO. 3. RECIPIENT'S CATAI	
FEASIBILITY OF USING FLUORESCENT MATER PRODUCT ASSURANCE APPLICATIONS AND FOR		PERIOD COVERED
ADHESIVE BOND FRACTURES	6. PERFORMING ORG.	REPORT HUMBER
AUTHOR(s)	8. CONTRACT OR GRA	NT NUMBER(a)
Carolyn A. L. Westerdahl J. Richard Hall	AMCMS Code 6121	.05.11H8.4
Performing organization name and address Feltman Research Laboratory Picatinny Arsenal Dover, NJ 07801	10. PROGRAM ELEMEN AREA & WORK UNIT	T, PROJECT, TASK I NUMBERS
1. CONTROLLING OFFICE NAME AND ADDRESS ARRADCOM	12. REPORT DATE	
ATTN: DRDAR-TSS	March 1978	
Dover, NJ 07801	13. NUMBER OF PAGES	
ARRADCOM, LCWSL Applied Sciences Division (DRDAR-LCA)		(of this report)
Dover, NJ 07801	150. DECLASSIFICATION SCHEDULE	N/DOWNGRADING
17. DISTRIBUTION STATEMENT (of the abstract entered in Bloc	k 20, if different from Report)	
8. SUPPLEMENTARY NOTES		
	ly by block number)	
Adhesive bonding, process control Quality control	Spectra, fluorescence Failure, cohesive Failure, adhesive Detection, UV	

bonds.

DD 1 JAN 73 1473 EDITION OF 1 NOV 65 IS OBSOLETE

UNCLASSIFIED

UNCLASSIFIED		
SECURITY CLASSIFICATION OF	THIS PAGE(When Data Entered)	
19. KEYWORDS (Con	ntinued)	
4-chloro-7-nitrober Adhesives Coatings Polymers	nzofurazan (NBD chloride)	
Pigments, fluoresce Brighteners	ent	
	5 -	
5		

ACKNOWLEDGMENTS

The authors are grateful to Mrs. M. C. Ross for supplying samples, J. R. Jarrell for providing equipment, and Dr. D. W. Levi for his valuable suggestions.

TABLE OF CONTENTS

	Page No.
Introduction	1
Discussion and Results	2
Survey Fluorescence Labeling of Polymers Containing Amino Groups	2 3
Experimental	4
Materials Instruments Sample Handling	4 9 10
Conclusions	10
References	11
Distribution List	23
Tables	
1 Fluorescence spectra of adhesives	13
2 Fluorescence spectra of structural polymers	17
3 Fluorescence spectra of fluorescent pigments	19
4 Fluorescence spectra of solvents	21

INTRODUCTION

Automation of assembly lines and the increasing prevalence of 100% inspection make necessary the detection of adhesives, surface primers, and other materials at certain stations of the assembly line. For example, the presence of adhesive used as a bonding agent should be verified before the bonded components are assembled in the end item. On the other hand, adhesive accidentally dropped on a component should be detected and removed before final assembly. Because these materials are often colorless or of a noncontrasting color, direct detection is difficult and indirect methods, such as electric eye detection of falling drops or monitoring of the reservoir level, are used. Such methods are not completely satisfactory because they are indirect and only verify that the liquid is leaving the reservoir without indicating where it is going.

Fluorescent detection is one of the more promising methods of direct detection. If the material is naturally fluorescent there is no problem. If it is not, fluorescent dye may be added. It is then illuminated with an ultraviolet or black light source and the emitted visible light is detected visually or with instrumentation. The detectors for visible light are sensitive and inexpensive. With fiber optic guides available for the UV and visible regions, it is possible to detect a substance in a congested space or otherwise inaccessible position. Many polymers are naturally fluorescent so that their presence at the correct location in an item can be detected efficiently by fluorescence.

One of the big advantages of fluorescent detection is low back-ground interference which results from the fact that the exciting illumination is often far removed in wavelength from the fluorescent light emitted by the substance. This method is also efficient for quick visual inspection, particularly if easily obtained UV filtering glasses are worn. In this way traces of adhesives remaining on broken bonded adherends can be seen. By scanning the surface for traces of adhesive it is possible to determine whether the fracture took place within the bulk of the adhesive (cohesive failure) or at the interface between the adhesive and the adherend (adhesive failure). One then knows whether to concentrate on surface treatment or on the physical properties of the adhesive to solve the adhesive failure problem.

This report is a survey of some of the common adhesives, coatings, polymeric solids, pigments, and solvents. The wavelengths of the exciting and emitted light are included, as well as an indication of strengths of peaks.

DISCUSSION AND RESULTS

Survey

The materials investigated included representatives of most of the common types of adhesives (Table 1), many of the common structural polymers (Table 2), a series of fluorescent pigments (Table 3), and some common solvents (Table 4). These materials were chosen to give a nucleus of information about common fluorescent materials so that the feasibility of using this method of quality control could be investigated conveniently. Many of these materials are naturally fluorescent or contain small amounts of fluorescent impurities. This fluorescence makes a tag which is suitable for following the location of the substance of interest. Trace amounts of fluorescent dyes and paint can be added to transparent substances to give an adequate response to UV light.

Fluorescence occurs when an electron in an atom or molecule is excited from a ground state to a higher electronic state by a photon. It loses some energy by a nonradiative transition, then decays back to the ground state. Because of the energy loss in the nonradiative transition (e.g., vibrational cascade), the energy of the emitted light is less than that of the exciting light, i.e., has a longer wavelength. The fact that the wavelengths of the exciting and emitted light can be widely different means that there is almost no interference between them and therefore there is a low background. Fluorescence spectroscopy is capable of good discrimination because only certain wavelengths will excite a molecule and when it is excited it will emit only certain wavelengths. This means that if one restricts the exciting light to a certain wavelength range and accepts only certain emitted wavelengths, only a small percentage of fluorescent compounds will be detected. This fact enables one to doubly discriminate against unwanted fluorescent substances and to be very specific in detecting the material of interest.

Practical applications have been made of the detection of fluorescent adhesive. The fluorescence of a black polyurethane rubber adhesive, PR 1520, has been used to detect its presence on a black neoprene substrate in the Lance system (Ref 1). Broken adhesive-bonded tensile specimens are routinely examined under UV light because many of the adhesives fluoresce and the small broken bits remaining on the faying surfaces are much more easily detected than under regular light. Besides the adhesives listed in Table 1, Tame 200, PB2 and EC 2214, used in the Durability Program (Ref 2, 3) have been used to study the distribution of adhesives on failed specimens. In the Rotating Band Program (Ref 4), EC 2290 was detected on Nylon 12.

In the AVSCOM Helicopter Repair Program (Ref 5), the presence of ADX 656.2 RT, Metlbond 1113, and EA 9628 on failed tensile specimens was detected. Fluorescent pigment has been added to Pettman cement (Ref 6) in an attempt to make it detectable on an assembly line. However, the high loading (50%) of iron oxide masks the added fluorescence. The amount of pigment which would be necessary for detection would make the cost of the cement excessive.

When using ultraviolet light, care must be taken to avoid over-exposure of eyes and skin. In an automated line this is no problem and in a manual situation, opaque gloves and specially tinted glasses provide adequate protection.

Fluorescence Labeling of Polymers Containing Amino Groups

The use of NBD chloride (4-chloro-7-nitrobenzo-2-oxa-1,3-diazole) to detect and determine amino acids, peptides and other compounds containing amino groups has been described (Ref 7 through 10) on the basis of the fluorescent properties of the reaction products. The method consists of treating the amino compound with NBD chloride (also known as 4-chloro-7-nitrobenzofurazan) in dilute solution for a short time. The NBD chloride is nonfluorescent; the N-derivatives are strongly fluorescent. NBD chloride also reacts with -SH and -OH groups to produce derivatives which, although fluorescent, are less stongly so than those of amino compounds.

$$O_2N - \bigcirc O_2N - O_2N - \bigcirc O_2N - O_2N - \bigcirc O_2N - O_2$$

To test the feasibility of introducing fluorescent groups into the structures of nonfluorescent polymers containing amino groups, a "soluble" nylon was treated with NBD chloride in 80/20 ethanol/water. The nylon, Elvamid 8061 (DuPont), is a 6-6 nylon partially substituted with methoxymethyl groups. The solution showed strong fluorescence at 538 nm upon excitation at 415 nm. Solutions of NBD chloride in methanol or 80/20 ethanol/water also showed fluorescence in the same region, but much weaker and the excitation peak was at 469 nm.

In an earlier experiment, a piece of 6-12 nylon of a suitable shape for insertion into the spectrophotofluorometer was immersed in a 1% solution of NBD chloride for $1\frac{1}{2}$ hours and rinsed. It exhibited

strong fluorescence at 520 nm; excitation, at about 460 nm. The NBD chloride solution was not fluorescent. (NBD chloride solutions prepared later did show some fluorescence. This appeared to be due either to slight contamination with amino vapors in the laboratory air or a slow reaction with -OH groups.) Untreated 6-12 nylon showed little or no fluorescence over the entire range.

The foregoing demonstrates the potential usefulness of NBD chloride as a fluorometric reagent for nylons and N-containing polymers. However, one of the investigators developed a rash from handling NBD chloride, indicating the need for care in the use of this compound.

EXPERIMENTAL

Materials

Solvents:

Styrene, stabilized, reagent grade.
2-methoxyethanol, scintillation grade.
Acetone, practical grade.
Isopropyl alcohol, reagent grade.
Heptane, chromatoquality grade.
Toluene, tech grade.
Ethanol, absolute, reagent grade.
Methanol, reagent grade.

Poly (4-methy1-1-pentene)

Polymers:

Poly(vinyl fluoride)

Tedlar, 200-SG40-TR,
E. I. du Pont de Nemours
and Company, Wilmington, DE

Poly(oxymethylene)

Celcon, M90-01, injectionmolding grade (MIL-A-50414),
Celanese Corp

TPX, ICI America, Inc

Nylon 6 Capran 77C, Allied Chemical Corp

Nylon 66 Almac Plastics, Long Island City, NY

Poly(ethylene terephthalate)

Mylar D, cartography grade, E. I. de Pont de Nemours and Company, Wilmington, DE

Cellulose acetate butyrate

film

Almac Plastics, Long Island

City, NY

Polycarbonate film

Lexan 8070-112, General

Electric

Polyethylene fluff

6050, Phillips Petroleum,

Bartlesville, OK

Polypropylene film

Dow 201, Dow Chemical,

Midland, MI

Nylon 12

Rilsan Industrial Inc,

Birdsboro, PA

Nylon 6-12

E. I. du Pont de Nemours,

Wilmington, DE

Nylon 11

Rilsan Industrial Inc,

Birdsboro, PA

Poly(chlorotrifluoro-

ethylene)

Kel-F, Adam Spence, Edison,

NJ

Poly(vinylidene) co-

polymer

Saran F-310 powder, Dow Chemical, Midland, MI

Adhesives:

Epon 828

Epoxy resin, diglycidyl.

ether of bisphenol A, Miller-

Stephenson Chem Co, Inc,

Danbury, CT

Accrabond 7521

Styrene monomer polyester,

Accrabond, Memphis, TN

PRC 1660

Polyurethane elastomer with

MEK (Parts A and B),

Products Research & Chemical

Corp, Burbank, CA

V-40	Polyamide resin, Miller- Stephenson Chem Co, Inc, Danbury, CT
EC 3549-B	3M Company, St Paul, MN
EC 2216-A, B	Epoxy adhesive: Part A, hardener-modified amine. Part B, base epoxy resin, 3M Company, St Paul, MN
RTV 118	Poly(dimethyl siloxane), Dow Corning Corp, Midland, MI
Flexcraft 1256-2	Synthetic resins in ketones and esters, Flexcraft Industries, Newark, NJ
Laminac 4116	Styrene monomer, American Cyanamid
Loctite 317	Anaerobic modified acrylic, Loctite Corporation, Newington, CT
Loctite 271	Anaerobic dimethacrylate, Loctite Corp, Newington, CT
Vibratite VC3	Nylok-Detroit, Troy, MI
Pettman Cement Type A	50% iron oxide, 20% ethyl alcohol, 12% pine tar, 18% shellac, Bradshaw Praeger & Co, Chicago, IL
Scotch Weld 2214	Aluminum-filled one-part- modified epoxy adhesive, 3M Company, St Paul, MN
Scotch Grip 1357	Polychloroprene in solvent, 3M Company, St Paul, MN

ADX 656.2

Supported epoxy, Hysol Division, Dexter Corp, Pittsburgh, CA

PL 717B	Supported epoxy, B. F. Goodrich General Products Company, Akron, OH
Metlbond 1113	Supported epoxy, Warmco Materials Inc, Costa Mesa, CA
EA 9628	Supported epoxy, Hysol Division, Dexter Corp, Pittsburgh, CA
Loctite 308	Anaerobic polyacrylate, Loctite Corporation, Newington, CT
Alfa 841	Aliphatic elastomeric polyurethane lacquer, Baker Castor Oil Co, Bayonne, NJ
Acryloid E 48-N 50/0	Acrylic resin, Rohm & Haas, Philadelphia, PA
EC 870	Oil resistant synthetic elastomer in toluol, 3M Company, St Paul, MN
Tame 200	Acrylic, B. F. Goodrich General Products Company, Akron, OH
EC 2214	Epoxy, 3M Company, St Paul, MN
PR 1520	Black polyurethane rubber, Products Research & Chemical Corp, Gloucester City, NJ
EC 833	Oil soluble elastomer in

0il soluble elastomer in petroleum naphtha, 3M Company, St Paul, MN

LP 2 Polymer of bis(ethylene-oxy)methane containing disulfide
linkages, terminated with reactive thiol groups, Thiokol,
Trenton, NJ

EC 1099	Nitrile rubber in ketone, 3M Company, St Paul, MN
Laminac 4134	Modified styrene, American Cyanamid Company, Wallingford, CT
Cascophen RS 216	Resorcinol in alcohol water, Borden Chemical, NY, NY
Penacolite	37% formaldehyde - water
Fluorescent Pigments:	
В 3556	Blue, Lawter Chemicals, Inc, Chicago, IL
В 3515	Gold yellow, Lawter Chemicals, Inc, Chicago, IL
В 3513	Red orange, Lawter Chemicals, Inc, Chicago, IL
В 3545	Green, Lawter Chemicals, Inc, Chicago, IL
В 3530	Cerise, Lawter Chemicals, Inc, Chicago, IL
B 3522	Pink, Lawter Chemicals, Inc, Chicago, IL
Uvitex OB	Fluorescent whitener, Ciba- Geigy Corp, Ardsley, NY
Tinopal PCRP	Fluorescent whitening agent, Ciba-Geigy Corp, Ardsley, NY
NBD Chloride	4-chloro-7-nitrobenzofurazan M.P. 97-99°C, Aldrich Chemical Company, Milwaukee, WI

Instruments

The spectrophotofluorometer, an Aminco Bowman instrument, consists of an optical unit, a photomultiplier microphotometer, an X-Y recorder, and accessory components. The optical unit includes a Xenon lamp, two monochrometers, slit holders, a cell compartment, and a photomultiplier housing. Light from the Xenon lamp is dispersed by the initial grating monochrometer into monochromatic radiation incident on the sample. The fluorescent radiation given off by the sample is dispersed by another grating into monochromatic radiation incident on the photomultiplier. The light produces a weak electrical signal which is fed to the photometer where it is amplified. This signal is sent to the recorder.

The gratings are operated by motor-driven cams coupled with graduated discs for visual observation and manual adjustment of wavelengths. One grating can be driven at a time. A potentiometer coupled to a grating supplies wavelength information to the recorder in the form of a DC voltage. Thus, with the exciting wavelength set, a graph of emission intensity versus wavelength may be obtained or, if the emission wavelength is set, a graph of intensity versus excitation wavelength is recorded.

The sample may be either liquid or solid. Liquids are usually placed in a 10 mm pathlength cuvette with four polished sides. The fluorescent emission is detected at 90° to the path of the exciting light. In a nonturbid solution, there is essentially no background from this light. There are also cylindrical microtubes with interior diameters of 2 mm. These can be used with strongly absorbing solutions, but tend to have more of a scattering background.

Finely divided solids can be used in either the cuvettes or tubes with some success. Strips or films can be placed in a special holder in which a mirror placed to the side of the exciting beam directs the light reflected from the sample into the emission monochrometer. This holder may also be used for opaque liquids. With this holder there is a large amount of background from the exciting light.

Slits can be inserted in the incoming beam and the emitted beam and before the photomultiplier to control the amount of light in the system and the bandwidth of the light.

The cuvettes are cleaned after each use by boiling them in concentrated nitric acid to eliminate any carryover of fluorescences.

Sample Handling

In running a sample, the sample was put into the cuvette which in turn was placed in the holder. A set of slits was selected, usually wide ones for the initial scans, and the microphotometer set to a moderate value. The excitation wavelength was set to 200 nm and the emission spectrum was run. If necessary, the sensitivity of the microphotometer was increased and the spectrum rerun. Then the excitation wavelength was increased to 250 nm and an emission spectrum was run. The excitation wavelength was increased in steps of 50 nm and an emission spectrum run for each. If there were any emission peaks, the emission wavelength was set on these and an excitation spectrum run. This was done for each emission peak. The spectra could be refined by using narrower slits as long as there was enough light.

Strongly absorbing substances were sometimes diluted with a non-absorbing solvent if a suitable one could be found. Several common solvents have large portions of their spectra free of absorption or emission peaks.

CONCLUSIONS

The use of UV light to excite fluorescence and thereby detect the presence of materials in their correct place on an assembly line is feasible and should be investigated when setting up 100% inspected criteria. Many organic materials are naturally fluorescent and most of the others can accept the presence of small amounts of a fluorescent pigment or label.

REFERENCES

- 1. M. D. Anderson and J. R. Hall, "Evaluation of Potting Problems in the LANCE AK Cable," Technical Report 4811, Picatinny Arsenal, Dover, NJ, 1975.
- M. C. Ross, W. C. Tanner, and M. J. Bodnar, "Effects of Varying Processing Parameters on the Fabrication of Adhesive Bonded Structures. XVII, Strength of Joints to Selected Corrosion-Resistant Finishes," Technical Report 4468, Picatinny Arsenal, Dover, NJ, 1973
- R. F. Wegman, M. C. Ross, E. A. Garnis, and S. A. Slota, "A Technique for Assessing the Durability of Structural Adhesives," ARRADCOM Technical Report, in press.
- 4. M. C. Ross, "Adhesive Bonding of Newer Structural Plastics," ARRADCOM Memorandum Report, in press.
- R. F. Wegman, M. C. Ross, E. A. Garnis, and S. A. Slota, "New Bonding Systems for Depot Maintenance of Aircraft Honeycomb Structures," Technical Report 4810, Picatinny Arsenal, Dover, NJ, November 1975.
- 6. M. D. Anderson, private communication, 1975.
- 7. P. B. Ghosh and M. W. Whitehouse, <u>Biochemical J</u>, <u>108</u>, 155-6 (1968).
- 8. J. Reisch, H.-J. Kommert, H. Alfes, and Z. Möllmann, <u>Anal Chem</u>, <u>247</u>, 56-7 (1969).
- 9. J. Reisch, H.-J. Kommert, and D. Clasing, <u>Pharm Ztg</u>, <u>115</u>, 752-3 (1970).
- 10. F. Rodier, M. Hill, B. Arrio, and C. Parquet, Fed Eur Biochem Soc Lett, 11, 246-8 (1970).

Table 1
Fluorescence spectra of adhesives*

Adhesive	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
Epon 828	<u><</u> 300	410 460 375-475 (VS)	360 385
Accrabond 7521	330 385 430	510 440 455 (S) 486 (S) 470	388 (W) 375 (S) 385 (S) 385 (S)
PRC 1660-L-A L-B	430-510 220-240 260-300	490-600 (S) 550 350-520 (VS) 325-480 (VS) 630 (M) 325 380 470	465 (S) 297 (M) 330 (VS) 440 (S), 360 (M)
EC 3549B	\[\leq \frac{360}{240 - 300} \\ \leq 200 - 360 \]	590 (VW) 350 (W) 425-520 (M) 400	300 (W)
EC 2216A	400 240-400	460 (VS) 450-500 (W) 460 500	400 + (W) 410-470 (M)
V-40	<u>></u> 200	510 (VS)	450 (VS)
EC 2216B	> 200 440-460	370 400 430 460 490 520 550 350-450 (VS) 500 (W)	330 (VS) 340 (VS) 355 (VS) 375 (VS) 387 (S) 493 (M) 400 (W)

Table 1 (Continued)

<u>Adhesive</u>	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
RTV 118	200-280	325-425 (W) 320 370	None None
Flexcraft 1256-2	200-300	325-375 (M) 332	None
Laminac 4116	300 - 340 320 - 400	370 (VS) 430 (VS) 440	367 (VS)
Loctite 317 (amber)	320+	475 (VS) 475	420 (S)
Loctite 271 (red) microcell	300-360 300	460 (VS) 450	355 (VS)
Vibratite VC 3	200-250	600 600 (VS)	350;470+ (W)
Pettman Cement Lot 2586	200-400	350-550 (W)	
Scotch Weld 2214	240-280	305 (S) 200	None
Scotch Grip 1357	200-510	None	
EC 2290 on Ny1on 11	200-350	365 to 430 (S) 365 450 510	310 (VW) 360 (W) 380 (W)
EC 2290 on Nylon 12	250-340 280-360	365 (W) 440 (W)	
A1413B on Nylon 11			
A1413B on Ny1on 12	350-400	440 (S)	

Table 1 (Continued)

Adhesive	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
ADX 6562 RT	250-350	330-400 (M) 350-500	None
PL 717B			
Metlbond 1113	250-350	390-460 (M) 420 430	275, 320 (W) 275, 335 (W)
EA 9628	200-350	335-470 (S) 350 400-450 500	No Sep Peak 320-60 (W) 385 (M)
Alfa 841	200-300 200-360	400-500 (W) 360-500 (S) 550 510 450 <400	400 (W) 380 (M) 358 (W) No Sep Peak
EC 870			
Loctite 308	350-500	450-550 (S) 450 530	395 (M) 447 (S)
LP-2	400-480	522 550-40 (VS)	455 (VS)
EC 833	250-400	410~490 (S) 455	380 (M)
Laminac 4134	320-450 (S) 365	400-500 (W) 450 (W) 500 460 440	380 (VS) 370 (VS) 365 (VS)
EC 1099 (Reflection)	250-350	380-450 (W) 400-425	290-350 (VW)

Table 1 (Continued)

Adhesive	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
EC 1099		61.0	5.5
(Microcell in acetone)	250-350	380-500 (S) 400-500 415	310, 360 (M) 310, 360 (W)
(Microcell in toluene)		415	310, 360 (W)
Cascophen R5216		380	310 (M)
	250-300	350-390 (M) 365	310 (S)
Penacolite G-1124B	200-250	305 360 280-380 (M)	220, 265 (W) 250-280 (W)

^{*}VS = Very stong fluorescence

VS and S would show easily detectable fluorescence.

S = Strong fluorescence

M = Moderate fluorescence

W = Weak fluorescence

VW = Very weak fluorescence

M would be detectable with some difficulty.

W and VW would not generally be usable.

Table 2
Fluorescence spectra of structural polymers*

Polymer	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
Celcon Film M9004	200-220	250 (M), 300-550 (S) 500 465	355 (M) 358 (M)
Tedlar	<u><</u> 230	250, 350, 500 (S) 503 468 350	210 (W) 200 (W) 220 (VW)
ТРХ	<210 <230 <250	465 (W) 260-300, 510 (W) 390 (W) 500	210 (W)
Polyethylene fluff 6050	223 (230) 230 (240+)	260 (M), 330-380 (S) 330 (S)-380 (W) 340, 230 (W)	
CAB	220-310	350 (M)	250, 285 (M)
TFE	<u><220</u>	250 (W), 337 (M) 485 (M) 460 485	320 (W) 300-400 (W)
	<u><210</u>	225, 462 (W) 337	220 (W)
Lexan Film	<260 ≤370	300 (M) 400 (M) 380	290 (M)
Polypropylene	200 220 <240	465;490 (M) 340, 526, 250 (M) 290 (W) 390 (W)	
		340	222 (W)

Table 2 (Continued)

<u>Polymer</u>	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
Nylon 12 30% glass filled	350-450 (385) 200-400	450 (S) 440, 470-570 (S)	
Nylon 6-12 33% glass	No peaks 350-500	500 (M-S) 490	380 (W)
Nylon 11	300-350	400 (W) 400 420 440 480 510	No peak 340 (W) 355 (W) 380 (W) 395 (W)
Nylon 6	200-350	400-470 (M) 450 410	350 (W) 340 (W)
Nylon 6-6	300-350	400 (W-M) 390 510	340 (W) 390 (W)
Kel F	None	None	
Mylar	200 225	260 (M), 525 (S) 260 (M), 480-560 (V	V −M)
Saran F-310	200-450	500-600 (530)(S) 570-530	450-225 (W)

^{*}VS = Very strong fluorescence

S = Strong fluorescence

M = Moderate fluorescence

W = Weak fluorescence

VW = Very weak fluorescence

VS and S would show easily detectable fluorescence.

M would be detectable with some difficulty.

W and VW would not generally be usable.

Table 3 Fluorescence spectra of fluorescent pigments*

Pigment	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitation wavelength (observed) nm
Blue Lawter B5556 in acetone	320-400 360 480	410-430 (M) 480 (M) 540 (W) 530 410-490	420 (M) 390-430 (M)
Gold Yellow Lawter 3515 in acetone	350-490	545-600 (W) 565	345 (M) 455 (S)
Red Orange B3513 in acetone	350-550	595 (VS) 592	330 (W) 355 (M-W) 460 (M)
Green B3545 in acetone	320-400	500 (S) 500	345 (VW) 460 (M)
Cerise B3530 in acetone	350-390	595 (VS) 595	350-465
Pink B3522 in acetone	300-500	588 (M) 590	300 (W) 345 (M) 400 (M) 475 (S)
Uvitex OB in toluene	300-400	435 (W) 420-450	280-340 (VS) 408 (VS)

Table 3 (Continued)

<u>Pigment</u>	Excitation wavelength, nm (Set)	Emission wavelength, nm	Excitiation wavelength (observed) nm
PCRP in toluene	310-400	400-440 (W) 420-460	300-350 (S) 385-400 (VS)

VS and S would show easily detectable fluorescence.

M would be detectable with some difficulty.

W and VW would not generally be usable.

^{*}VS = Very strong fluorescence

S = Strong fluorescence

M = Moderate fluorescence

W = Weak fluorescence

VW = Very weak fluorescence

Table 4
Fluorescence spectra of solvents*

Solvent	Excitation wavelengths, nm (Set)	Emission wavelengths, nm	Excitation wavelengths (observed) nm
Styrene	300-340	350-410 (S) 385	340 (M)
Acetone	260-320 300-340	335 (M) 385 (M) 390 330	333 (M) None
Isopropy1 Alcohol (IPA)	220-300 200-260	340-440 (M) 290-320 (W) 390 (W)	290 (W)
н ₂ 0	220-300	310-340 (VW)	
90% IPA + H ₂ 0	210-270 210-270	290-310 (W) 320-360 (M) 340	225 (W)
Heptane	200-300 260-300 220	340 (S) 610-650 (W) 510 (W) 330 625	280 (S) 280 (M)
Toluene	250-350 282	280-410 (M) 330-410 330 (S)	280-290 (M)
Abs Ethyl Alcohol	200-300	290-350 (M) 310 310	215 (W) 275 (M)
Methyl Alcohol	200-250 250-350 700-350	295 (S) 350-420 (M) 540-570 (M)	ð

Table 4 (Continued)

Excitation wavelengths, mm (Set) mm Excitation wavelengths, nm (Set) nm (Observed) nm

2 Methoxyethanol 200-300 600 (W)

VS and S would show easily detectable fluorescence. M would be detectable with some difficulty. W and VW would not generally be usable.

^{*}VS = Very strong fluorescence

S = Strong fluorescence

M = Moderate fluorescence

W = Weak fluorescence

VW = Very weak fluorescence

DISTRIBUTION LIST

	Copy No.
Commander	
US Army Armament Research & Development Command	
ATTN: DRDAR-TSS	1-5
DRDAR-LCA-OA	6-20
DRDAR-LCA-OK, Mr. Pebly	21
Mr. Landrock	22
DRDAR-LCN	23-27
DRDAR-LCV	28-32
DRDAR-QA	33-34
DRDAR-TSF	35-36
DRDAR-QAA	37
DRDAR-QAN	38
Dover, NJ 07801	50
DOVE1, NO 07601	
Commander	
US Army Materiel Development & Readiness Command	
ATTN: DRCDE-R, Mr. J. Rivkin	39
DRCPP-PI	40
DRCQA	41
5001 Eisenhower Avenue	
Alexandria, VA 22304	
Commander	
US Army Missile Research & Development Command	
ATTN: DRSMI-RLM, Mr. E. A. Verchot	42
Chief, Document Section	43
Redstone Arsenal, AL 35801	
Commander	
US Army Armament Materiel Readiness Command	
ATTN: DRSAR-MP-PC	44
DRSAR-RDP	45
SARRI-SP	46
DRSAR-RDM	47
DRSAR-RDT, Dr. Daryl Penrod	48
DRSAR-ASF, Mr. H. Wohlferth	49
Pook Island II 61201	

Commander	
US Army Electronics Command	
ATTN: DRSEL-TL-ME, Mr. Dan Lichenstein	50
DRSEL-TL-ME, Mr. A. J. Raffalovich	51
DRSEL-TL-ME, Mr. G. Platau	52
DRSEL-PP-EM2, Sarah Rosen	53
Fort Monmouth, NJ 07703	33
Commander	
US Army Aviation Systems Command	
ATTN: DRSAV-FET, Mr. J. Bramlet	54
DRSAV-EER, Mr. W. McClane	55
DRSAV-FEU, Mr. E. Dawson	56
DRSAV-FFE, Mr. C. Sims	57
DRSAV-REN, Mr. L. D. Brown	58
P.O. Box 209, Main Office	30
St Louis, MO 63166	
,	
Director	
US Army Tank-Automotive Research and	
Development Command	
ATTN: DRSTRA-KMD, Mr. Francis S. Lemmer	59
Warren, MI 48090	
Commander	
US Army Materials & Mechanics Research Center	
ATTN: DRXMR-FR, Dr. G. Thomas	60
DRXMR-FL	61
Technical Information Section	62
Watertown, MA 02172	-
Director	
US Army Production Equipment Agency	
Rock Island Arsenal	
ATTN: DRXPE-MT, Mr. H. Holmes	63-64
Rock Island Arsenal, IL 61201	05 04
,	
Commander	
Corpus Christi Army Depot	
ATTN: DRSAV-FES (STOP 55)	65-66
DRSAV-FESA, Mr. T. Tullos	67-68
DRSAV-FESP, Mr. Bulloch	69
Cornus Christi TY 78410	U J

Commander/Director Chemical Systems Laboratory	70
US Army Armament Research and Development	
Command, Bldg E5101	
Aberdeen Proving Ground, MD 21010	
Chief	
Benet Weapons Laboratory	
LCWSL, USA ARRADCOM	71
ATTN: DRDAR-LCB Watervliet, NY 12189	/1
watervilet, Ni 12109	
Director	
US Army Engineer Waterways	72
ATTN: Mr. Hugh L. Green - WE SSS1 Experiment Station, P.O. Box 631	12
Corps of Engineers	
Vicksburg, MS 39180	
vicksburg, no ostoo	
Commander	
US Army Medical Bio-Engineering Research	
and Development Laboratories	
Fort Deterick	
ATTN: Dr. C. Wade	73
Frederick, MD 21701	
Commander	
Harry Diamond Laboratories	
ATTN: Mr. N. Kaplan	74
Mr. J. M. Boyd	75
Library	76
Washington, DC 20438	
Commander	
US Army Armament Research and Development Command	
ATTN: DRDAR-CSL, Mr. Dave Schneck	77
Aberdeen Proving Ground, MD 21010	
Commander	
Tobyhanna Army Depot	
ATTN: Mr. A. Alfano	78
Tobyhanna, PA 18466	
Dimenton	70
Director US Army Ballistic Research Laboratory	79
USA ARRADCOM, Bldg 328	
Aberdeen Proving Ground, MD 21005	
, , , , , , , , , , , , , , , , , , , ,	

Commander	
US Army Materiel Development & Readiness Command	
ATTN: DRCPM-UA, Mr. C. Musgrave	80
DRCPM-LH, Mr. C. Cioffi	81
DRCPM-HLS-T, Mr. R. E. Hahn	82
P.O. Box 209	
St Louis, MO 63166	
Commander	83
Natick Research and Development Command	
Natick, MA 01760	
Commander	84
US Army Engineer Research & Development Labs	
Fort Belvoir, VA 22060	
D	
Department of the Navy	
Naval Air Systems Command	· 0 E
ATTN: Mr. John J. Gurtowski (AIR 520320)	85
Washington, DC 20360	
Naval Ordnance Station (NOSL)	
ATTN: Mr. W. J. Ryan Code 5041	86
Southside Drive	00
Louisville, KY 40214	
hoursville, ki 40214	
Naval Avionics Facility	
ATTN: Mr. B. D. Tague, Code D/802	87
Mr. Paul H. Guhl, D/033.3	88
21st and Arlington	
Indianapolis, IN 46218	
Commander	
US Naval Weapons Station	
ATTN: Research and Development Division	89
Yorktown, VA 23491	
Commander	
Aeronautical Systems Division	
ATTN: Mr. W. Scardino, AFML/MXE	90
Mr. T. J. Aponyl	91
Composite and Fibrous Materials Branch	
Nonmetallic Materials Division	
Wright-Patterson Air Force Base, OH 45433	

Dr. Robert S. Shane, Staff Scientist National Materials Advisory Board National Academy of Sciences 2101 Constitution Avenue, NW Washington, DC 20418	92
US Army Air Mobility R&D Laboratory, Headquarters Advanced Systems Research Office ATTN: Mr. F. Immen, MS 207.5 Ames Research Center Moffet Field, CA 94035	93
Naval Ship Engineering Center ATTN: Mr. W. R. Graner, SEC 6101E Prince George's Center Hyattsville, MD 20782	94
Mare Island Naval Shipyard ATTN: Mr. Ross E. Morris, Code 134.04 Rubber Engineering Section Vallejo, CA 94592	95
Hanscom Air Force Base ATTN: Mr. R. Karlson, ESD/DE, Stop 7 HQ, ESD Bedford, MA 01731	96
Naval Air Development Center	
Materials Laboratory ATTN: Mr. Coleman Nadler, Code 30221, Div, AVTD Warminster, PA 18974	97
Defense Documentation Center Cameron Station Alexandria, VA 22314	98-109
Weapon System Concept Team/CSL ATTN: DRDAR-ACW Aberdeen Proving Ground, MD 21010	110
Technical Library ATTN: DRDAR-CLJ-L Aberdeen Proving Cround MD 21010	111

Technical Library	
ATTN: DRDAR-TSB-S	112
Aberdeen Proving Ground, MD 21005	
Benet Weapons Laboratory	
Technical Library	
ATTN: DRDAR-LCB-TL	113
Watervliet, NY 12189	
Commander	
US Army Armament Materiel and Readiness Command	
ATTN: DRSAR-LEP-L	114
Rock Island, IL 61299	
director	
US Army TRADOC Systems Analysis Activity	
ATTN: ATAA-SL (Technical Library)	115
White Sands Missile Range, NM 88002	•